

Article

A Highly Selective Fluorescent Chemosensor for Pb

Ji Young Kwon, Yun Jung Jang, Yoon Ju Lee, Kwan Mook Kim, Mi Sook Seo, Wonwoo Nam, and Juyoung Yoon

J. Am. Chem. Soc., 2005, 127 (28), 10107-10111• DOI: 10.1021/ja051075b • Publication Date (Web): 25 June 2005

Downloaded from http://pubs.acs.org on March 25, 2009



More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Links to the 46 articles that cite this article, as of the time of this article download
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

View the Full Text HTML





A Highly Selective Fluorescent Chemosensor for Pb²⁺

Ji Young Kwon,[†] Yun Jung Jang,[†] Yoon Ju Lee,[†] Kwan Mook Kim,^{†,‡} Mi Sook Seo,^{†,‡} Wonwoo Nam,^{*,†,‡} and Juyoung Yoon^{*,†}

Contribution from the Department of Chemistry and Division of Nano Sciences, Ewha Womans University, 11-1 Daehyon-Dong, Sodaemun-Ku, Seoul 120-750, Korea, and Creative Research Initiative Program for Biomimetic Systems, Ewha Womans University, 11-1 Daehyon-Dong, Sodaemun-Ku, Seoul 120-750, Korea

Received March 2, 2005; E-mail: jyoon@ewha.ac.kr; wwnam@ewha.ac.kr

Abstract: A new fluorescent sensor based on rhodamine B for Pb²⁺ was synthesized. The new fluorescent sensor showed an extreme selectivity for Pb2+ over other metal ions examined in acetonitrile. Upon the addition of Pb2+, an overall emission change of 100-fold was observed, and the selectivity was calculated to be 200 times that of Zn²⁺. The signal transduction occurs via of reversible CHEF (chelation-enhanced fluorescence) with this inherent quenching metal ion.

Introduction

The selective binding of chemical species upon molecular recognition can lead to large perturbations in the host environment, particularly when the guest is ionic. Since fluoroionophores can provide chemical information on the ion concentrations, they are important subjects in metal ion analysis.¹

Among the metal ions, Pb²⁺ is one of the important targets because of the adverse health effects of lead exposure, particularly in children.² Even though considerable efforts have been devoted to developing fluorescent chemosensors for various metal ions over the last few decades,¹ there have been relatively few reports on Pb²⁺-selective fluorescent chemosensors. Lu et al. utilized catalytic DNAs as a unique class of biosensors for Pb^{2+.3} Ma and co-workers reported a fluorescent probe based on the Pb²⁺-catalyzed hydrolysis of phosphodiester.⁴ Even though the above results were excellent examples of Pb^{2+} selective fluorescent sensors, the probes were hydrolyzed by Pb²⁺ in both cases, and these are irreversible processes. For a real-time fluorescent sensor, Chen et al. used a ketoaminocoumarine derivative bearing a azacrown binding site as a new ratiometric fluorescent sensor for Pb2+.5 Godwin et al. also reported a new ratiometric fluorescent sensor for Pb2+ based

on the dansyl-tetrapeptide framework.⁶ On the other hand, Czarnik and co-workers reported a N-methyl-9-anthrylthiohydroxamic acid as a new fluorescent chemosensor for Pb2+.7 In addition, some of fluorophore-appended macrocycles have been reported.8

Rhodamine dyes have been used extensively for conjugation with biomolecules, owing to their excellent fluorescence properties. A few rhodamine B derivatives have also been used as fluorescent chemosensors for metal ions.9,10 Czarnik et al. reported that rhodamine B hydrazide could be used as a fluorescent chemodosimeter for Cu2+.9

We report a new fluorescent sensor 1 based on rhodamine B for Pb^{2+} . The structure of compound **1** was confirmed by X-ray crystallography in addition to NMR and mass data. The new fluorescent sensor 1 displayed an extreme selectivity for Pb²⁺ in acetonitrile compared with other metal ions examined. The signal transduction occurs via of reversible CHEF (chelationenhanced fluorescence) with this inherent quenching metal ion. ¹H NMR, ¹³C NMR, IR, and the electrospary ionization (ESI) mass data were used to explain the binding mode of 1 with metal ions.

Experimental Section

General Methods. Unless otherwise noted, materials were obtained from commercial suppliers and were used without further purification.

[†] Department of Chemistry and Division of Nano Sciences. [‡] Creative Research Initiative Program for Biomimetic Systems.

^{(1) (}a) Fluorescent Chemosensors for Ion and Molecular Recognition; Czarnik, (a) Fulorescent Chemosensors for 1on and Molecular Recognition; Czarnik, A. W., Ed.; American Chemical Society: Washington, DC, 1993. (b) Czarnik, A. W. Acc. Chem. Res. 1994, 27, 302. (c) Fabbrizzi, L.; Poggi, A. Chem. Soc. Rev. 1994, 197. (d) de Silva, A. P.; Gunaratne, H. Q. N.; Gunnlaugsson, T. A.; Huxley, T. M.; McCoy, C. P.; Rademacher, J. T.; Rice, T. E. Chem. Rev. 1997, 97, 1515. (e) Chemosensors of Ion and Molecular Recognition; Desvergne, J.-P., Czarnik, A. W., Eds.; Kluwer Academic Dublicheru, Deordersch, The Nithorleade, 1007. (f) Amendel. Academic Publishers: Dordrecht, The Netherlands, 1997. (f) Amendola, V.; Fabbrizzi, L.; Lincchelli, M.; Mangano, C.; Pallavicini, P.; Parodi, L.; Poggi, A. *Coord. Chem. Chem. Rev.* **1999**, *190–192*, 649.

⁽²⁾ Needleman, H. L. Human Lead Exposure; CRC Press: Boca Raton, FL, 1992.

^{(3) (}a) Liu, J.; Lu, Y. J. Am. Chem. Soc. 2003, 125, 6642. (b) Li, J.; Lu, Y. J.

 ⁽³⁾ Lu, J., Eu, T. J. Am. Chem. Soc. 2009, 123, 0042. (b) El, J., Eu, T. J. Am. Chem. Soc. 2000, 122, 10466.
(4) Sun, M.; Shangguan, D.; Ma, H.; Nie, L.; Li, X.; Xiong, S.; Liu, G.; Thiemann, W. Biopolymers 2003, 72, 413.
(5) Chen, C.-T.; Huang, W.-P. J. Am. Chem. Soc. 2002, 124, 6246.

⁽⁶⁾ Deo, S.; Godwin, H. A. J. Am. Chem. Soc. 2000, 122, 174

 ⁽⁶⁾ Deo, S., Gouvin, H. A. J. Am. Chem. Soc. 2000, 122, 174.
(7) Chae, M.-Y.; Yoon, J.; Czarnik, A. W. J. Mol. Recognit. 1996, 9, 297.
(8) (a) Xia, W.-S.; Schmehl, R. H.; Li, C.-J.; Mague, J. T.; Iuo, C.-P.; Guldi, D. M. J. Phy. Chem. B 2002, 106, 833. (b) Padilla-Tosta, M. E.; Lloris, J. M.; Martinez-Máñez, R.; Marcos, M. D.; Miranda, M. A.; Pardo, T.; Sancenón, F.; Soto, J. Eur. J. Inorg. Chem. 2001, 1475. (c) Addleman, R. S.; Parentt, J.; Tungdy, S. H.; Eldoman, S. H.; Eldoman, S.; M.; C. M. T., and S. H.; Eldoman, R. S.; Parentt, J.; Tungdy, S. H.; Eldoman, R.; Superst, J. & Tungdy, S. H.; Eldoman, R. S.; Parentt, J.; Tungdy, S. H.; Eldoman, R.; Superst, J. S.; M.; Marcos, M. B.; J. Superst, J. S.; M.; Marcos, M. B.; J. Superst, J. S.; M.; Eldoman, R.; Superst, J. S.; M.; Marcos, M.; Marcos, S.; M.; Eldoman, R.; Superst, J. S.; M.; Eldoman, R.; Superst, J. S.; M.; Eldoman, R.; Superst, J. S.; M.; Marcos, M.; Marcos, S.; M.; Eldoman, R.; Superst, J.; Superst, J.; J.; J.; M.; Marcos, M.; M.; Marcos, S.; M.; Eldoman, R.; Superst, J.; Superst S.; Bennett, J.; Tweedy, S. H.; Elshani, S.; Wai, C. M. *Talanta* 1998, 46, 573. (d) Shen, Y.; Sullivan, B. P. *J. Chem. Educ.* 1997, 74, 685. (e) Beeby, A.; Parker, D.; Williams, J. A. G. *J. Chem. Soc., Perkin Trans.* 2 1996, 1565.

^{(9) (}a) Dujols, V.; Ford, F.; Czarnik, A. W. J. Am. Chem. Soc. 1997, 119, 7386.

^{(10) (}a) Hojo, M.; Ueda, T.; Yamasaki, M.; Inoue, A.; Tokita, S.; Yanagita, M. Bull. Chem. Soc. Jpn. 2002, 75, 1569. (b) Thorn, D. L.; Fultz, W. C. J. Phys. Chem. 1989, 96, 1234. (c) Arbeloa, L.; Rohatgi-Mukherjee, K. K. Chem. Phys. Lett. 1986, 128, 474. (d) Oshima, G.-I.; Nagasawa, K. Chem. Pharm. Bull. 1970, 18, 687.

Flash chromatography was carried out on silica gel 60 (230–400 mesh ASTM; Merck). Thin-layer chromatography (TLC) was carried out using Merck 60 F_{254} plates with a thickness of 0.25 mm. Preparative TLC was performed using Merck 60 F_{254} plates with a thickness of 1 mm.

Melting points were measured using a Buchi 530 melting point apparatus, and are uncorrected. ¹H NMR and ¹³C NMR spectra were recorded using Bruker 250 or Varian 500. Chemical shifts were expressed in ppm and coupling constants (*J*) in Hz. Mass spectra were obtained using a JMS-HX 110A/110A Tandem Mass Spectrometer (JEOL). UV absorption spectra were obtained on UVIKON 933 Double Beam UV/vis Spectrometer. Fluorescence emission spectra were obtained using RF-5301/PC Spectrofluorophotometer (Shimadzu). Electrospray ionization mass (ESI-MS) spectra were performed on a Thermo Finnigan (San Jose, CA) LCQ Advantage MAX quadrupole ion trap instrument with samples of metal complexes.

1 (RBDPA-1). A solution of rhodamine B base (1.0 g, 2.3 mmol) in 1,2 dichloromethane (12 mL) was stirred, and phosphorus oxychloride (0.6 mL) was added dropwise over 5 min. The solution was refluxed for 4 h. The reaction mixture was cooled and evaporated in vacuo to give rhodamine B acid chloride, which was not purified but confirmed with the reported ¹H NMR.⁹

The crude acid chloride was dissolved in acetonitrile (125 mL) and added dropwise over 5 h to a solution of (2-aminoethyl)bis(2-pyridylmethyl)amine¹¹ (1.1 g, 4.6 mmol) in acetonitrile (50 mL) at room temperature. The reaction mixture was then refluxed for 1 h. After the solvent was evaporated under reduced pressure, the crude product was purified by column chromatography (CHCl₃:MeOH = 9:1, v/v) to give the 643 mg of **1** (yield; 42%): mp 56–59 °C.; ¹H NMR (CDCl₃) δ 8.44 (d, 2H, J = 4.8 Hz), 7.87 (t, 1H, J = 2.9 Hz), 7.57 (t, 1H, J = 7.7 Hz), 7.41 (m, 4H), 7.06 (m, 3H), 6.35 (m, 4H), 6.20 (dd, 2H, J = 8.8 and 2.6 Hz), 3.69 (s, 4H), 3.34 (m, 10H), 2.38 (t, 2H, J = 7.9 Hz), 1.16 (t, 2H, J = 7.0 Hz);¹³C NMR (CDCl₃) δ 168.4, 159.3, 153.9, 153.6, 149.8, 137.5, 132.7, 131.6, 129.2, 128.4, 126.3, 124.2, 123.4, 123.1, 122.3, 108.6, 108.4, 105.8, 98.3, 65.3, 60.1, 52.6, 44.8, 38.6, 14.5; HRMS (FAB) m/z = 667.3755 (M + H)⁺, calcd for C₄₂H₄₇N₆O₂ = 667.3761.

Preparation of Fluorometric Metal Ion Titration Solutions. Stock solutions (1 mM) of the perchlorate salts of Ag⁺, Ca²⁺, Cd²⁺, Co²⁺, Cs⁺, Cu²⁺, Hg²⁺, K⁺, Li⁺, Mg²⁺, Mn²⁺, Na⁺, Ni²⁺, Pb²⁺, Rb⁺ and Zn²⁺ in acetonitrile were prepared. Stock solution of host (0.1 mM) was also prepared in acetonitrile. Test solutions were prepared by placing 4–40 μ L of the probe stock solution into a test tube, adding an appropriate aliquot of each metal stock, and diluting the solution to 4 mL with acetonitrile.

For all measurements, excitation wavelength was 510 nm. Both excitation and emission slit widths were 5 nm.

Electrospray Ionization Mass (ESI-MS). The electrospray ionization mass (ESI-MS) spectrum of **1** was obtained by direct introduction of the solution of samples (10 μ M) in CH₃CN into the source at 25 μ L/min using a syringe pump. The spray voltage of the spectrometer was set at 5 kV, and the capillary temperature, at 150 °C.

X-ray Data. All the X-ray data were collected on a SMART APEX CCD equipped with a Mo X-ray tube at the ambient temperature. Crystal data of 1: *monoclinic*, P_{1} (No. 4), Z = 2, a = 11.896(3) Å, b = 11.812(3) Å, and c = 12.988(3) Å, $\beta = 90.592(4)^{\circ}$, V = 1824.8-(7) Å³, $\mu = 0.076$ mm⁻¹, $d_{calc} = 1.214$ g/cm³, R1 = 5.81% and wR2 = 11.62% for 1946 unique reflections and 451 variables. The structure solution and refinement of the data were handled with the SHELXS-86 and SHELXL-97 programs.

Results and Discussion

(2-Aminoethyl)bis(2-pyridylmethyl)amine was synthesized according to the published procedure.¹¹ $\mathbf{1}$ was synthesized by



treating rhodamine B with $POCl_3$,^{4a} which was followed without purification by (2-aminoethyl)bis(2-pyridylmethyl)amine. After column chromatography using CHCl₃:MeOH (9:1, v/v), **1** was obtained in a 42% yield (Scheme 1).

A single crystal of **1** was grown from a CHCl₃/CH₃CN solution and was characterized using X-ray crystallography (Figure 1). The crystal structure clearly represents the unique spirolactam-ring formation. Even though the crystal and molecular structures of several fluoran-based color formers by single-crystal X-ray diffraction analysis have been reported,¹² as far as we are aware, the X-ray structure of the rhodamine B derivative bearing the lactam moiety has not been reported.

The perchlorate salts of Ag^+ , Ca^{2+} , Cd^{2+} , Co^{2+} , Cs^+ , Cu^{2+} , Hg^{2+} , K^+ , Li^+ , Mg^{2+} , Mn^{2+} , Na^+ , Ni^{2+} , Pb^{2+} , Rb^+ , and Zn^{2+} ions were used to evaluate the metal ion binding properties of **1**. Stock solutions (1 mM) of the perchlorate salts of the metal ions in acetonitrile were prepared. A stock solution of the host (0.1 mM) in acetonitrile was also prepared. The test solutions were prepared by placing $4-40 \ \mu$ L of the probe stock solution into a test tube, adding the appropriate aliquot of each metal stock, and diluting the solution to 4 mL with acetonitrile. For all the measurements, excitation wavelength was 510 nm, and both the excitation and emission slit widths were 5 nm.

All titration studies were conducted using a 1 μ M concentration of **1**. Using these metal ions (100 equiv), **1** showed a large CHEF effect only with Pb²⁺, even though there was a relatively small CHEQ effect with Cu^{2+} and Zn^{2+} (Figure 2). Figure 3 shows the fluorescent emission changes of 1 upon the addition of Pb^{2+} ions. As shown in Figures 2 and 3, the addition of Pb^{2+} and Zn^{2+} caused a significant red shift (~30 nm). From the fluorescent titrations, the association constants for Pb²⁺ and Zn²⁺ were calculated to be 195000 and 900 M⁻¹, respectively (errors <10%).¹³ The selectivity for Pb²⁺ over Zn²⁺ was more than over 200 times. An overall emission change of 100-fold was observed for Pb²⁺. Furthermore, there was no significant change in the association constant for Pb^{2+} when excess Zn^{2+} (0.1 mM) was present. On the other hand, upon the addition of Cu^{2+} , 1 displayed a CHEF effect without a red shift. As shown in Figure 2, Li⁺ also caused a similar but smaller fluorescence change to Cu^{2+} .

⁽¹¹⁾ Schatz, M.; Leibold, M.; Foxon, S. P.; Weitzer, M.; Heinemann, F. W.; Hampel, F.; Walter, O.; Schindler, S. Dalton Trans. 2003, 1480.

^{(12) (}a) Okada, K. J. Mol. Struct. 1996, 380, 235. (b) Ribs, G.; Weis, C. D. Dyes Pigm. 1991, 15, 107 and 165.

^{(13) (}a) Conners, K. A. Binding Constants; Wiley: New York, 1987. (b) Association constants were obtained using the computer program ENZFTTER, available from Elsevier-BIOSOFT, 68 Hills Road, Cambridge CB2 ILA, United Kingdom.



Figure 1. Side and top view of X-ray crystal structure of 1.



Figure 2. Fluorescent emission changes of $1 (1 \mu M)$ upon addition of Ag⁺, Ca²⁺, Cd²⁺, Co²⁺, Cs⁺, Cu²⁺, Hg²⁺, K⁺, Li⁺, Mg²⁺, Mn²⁺, Na⁺, Ni²⁺, Pb²⁺, Rb⁺, and Zn²⁺ (100 equiv) in acetonitrile (excitation at 510 nm).



Figure 3. Fluorescent titrations of 1 (1 μ M) upon addition of Pb²⁺ in acetonitrile (excitation at 510 nm).

There was also a large enhancement (~120-fold) in the UV absorption ($\lambda_{\text{max}} = 553 \text{ nm}$) of **1** upon the addition of Pb²⁺, as shown in Figure 4. The fluorescence quantum yield was





Figure 4. UV absorption changes of 1 (0.1 mM) upon addition of Pb²⁺ in acetonitrile.

calculated as 52% by using perylene as a reference.¹⁴ 1 displayed a similar UV enhancement (\sim 20-fold) when Zn²⁺ was added to acetonitrile. On the other hand, when Cu²⁺ was added, the 1 did not display a new peak around 550 nm, and only broad absorption increase was observed around 450 nm (see Supporting Information).

Figure 5 shows the partial ¹H NMR spectra of **1** upon the addition of $Zn(ClO_4)_2$ (2 equiv). The NMR peaks were assigned on the basis of the ¹H-¹H COSY spectrum (see Supporting Information).

For example, upon the addition of 2 equiv of Zn^{2+} , the H_a and H_c on the pyridine ring in **1** showed relatively large downfield shifts (H_a , $\Delta \delta = 0.44$ ppm; H_c , $\Delta \delta = 0.54$ ppm).

The carbonyl carbon peak in the ¹³C NMR spectra moved from 167.5 to 171.5 ppm in CD₃CN:CDCl₃ (9:1, v/v) when 1.2 equiv of Zn(ClO₄)₂ was added (see Supporting Information). The cleavage of the lactone ring in the fluoran-based dye was reported and explained on the basis of the changes in the ¹³C NMR spectrum.^{10a,14} In the case of the fluoran-based dye, the spiro carbon in the lactone ring moiety appeared at approximately δ 75–85 ppm and moved to δ ~160 ppm in the

⁽¹⁴⁾ Cheng, Y.; Ma, B.; Wudl, F. J. Mater. Chem. 1999, 9, 2183.



Figure 5. Partial ¹H NMR (250 MHz) of 1 (2 mM) in CD₃CN. (a) 1 only; (b) 1 + 2 equiv of zinc(II) perchlorate.



Figure 6. Infrared spectra of 1 (a), 1 with $Zn(ClO_4)_2$ (b), 1 with $Pb(ClO_4)_2$ (c), and 1 with $Cu(ClO_4)_2$ (d).



Figure 7. ESI mass spectra of 1 with $Pb(ClO_4)_2$ (a) and 1 with $Zn(ClO_4)_2$ (b).

lactone ring-opened structure.^{10a,15} The spiro carbon in **1** appeared at δ 64.7 ppm in CD₃CN:CDCl₃ (9:1, v/v). When 1.2 equiv of Zn(ClO₄)₂ was added, the spiro carbon at δ 64.7 ppm disappeared and moved to δ 150 ppm. The addition of Pb(ClO₄)₂ caused a broadening of some peaks in the ¹H and ¹³C NMR spectra. However, the disappearance of the spiro carbon at δ 64.7 ppm was also confirmed. On the other hand, when 2.0 equiv of LiClO₄ was added, the spiro carbon peak was clearly observed at δ 66.6 ppm, and the carbonyl carbon peak moved from 167.5 to 170.5 ppm.

Figure 6 shows the partial IR spectra of 1 (50 mM) with 1 equiv of $Zn(ClO_4)_2$, Pb(ClO_4)₂, and Cu(ClO_4)₂ in CH₃CN-

CHCl₃ (4:1, v/v). The peak at 1689.53, which corresponds to the characteristic amide carbonyl absorption, was shifted to 1592.35 (with Zn^{2+}), 1614.72 (with Pb^{2+}), 1614.27 (with Cu^{2+}), and 1614.72 (with Li^+ , see Supporting Information). These results also support that a strong binding participation of the carbonyl group occurs with metal ions.

In the ESI mass spectrum of **1**, a peak at m/z 973.2 (calculated value, 973.3) corresponding to $[\mathbf{1} + \text{Pb}(\text{CIO}_4)]^+$ was clearly observed upon the addition of lead(II) perchlorate (Figure 7). In addition, a peak at m/z 437.1 (calculated value, 437.2) corresponding to $[\mathbf{1} + \text{Pb}]^{2+}$ was observed. Upon the addition of zinc(II) perchlorate, two distinct peaks at m/z 829.2 and 365.3 were observed, which correspond to $[\mathbf{1} + \text{Zn}(\text{CIO}_4)]^+$ and $[\mathbf{1}$

⁽¹⁵⁾ Yanagita, M.; Aoki, I.; Tokita, S. Bull. Chem. Soc. Jpn. 1997, 70, 2757.

Scheme 2. Proposed Mechanism for the Fluorescent Changes of 1 upon the Addition of Pb²⁺



+ Zn]²⁺, respectively. In a similar way, two distinct peaks at m/z 828.1 and 364.5 were observed with Cu²⁺, which correspond to $[1 + Cu(ClO_4)]^+$ and $[1 + Cu]^{2+}$, respectively (see Supporting Information). When cupper nitrate was added, a peak at m/z 791.2 (calculated value, 791.4) corresponding to $[1 + Cu(NO_3)]^+$ was observed (see Supporting Information).

The proposed mechanism for these fluorescent changes is explained in Scheme 2. Upon the addition of Pb²⁺ to a colorless solution of1 in acetonitrile, both a pink color and the fluorescence characteristics of rhodamine B appear. Because both disappear upon the addition of excess cyclen or ethylenediamine, it is believed that this process is reversible. Also, the ¹H NMR of the extracted product was identical to that of 1. The ¹³C NMR data also supports this spiro ring-opening mechanism. On the other hand, the addition of Cu2+ as well as some other metal ions caused relatively small CHEF effects without a red shift. This was also a reversible process since there was no change on TLC, and the characteristic fluorescent emission was recovered upon the addition of either cyclen or ethylenediamine. Furthermore, a chloroform and EDTA aqueous solution was added to the $1 \cdot Cu^{2+}$, and the ¹H NMR of the extracted product was identical to that of 1. As shown in Figure 2, the addition of Cu²⁺ and Li⁺ to 1 caused similar intensity changes. From the fact that the spiro carbon peak was still observed when 2.0 equiv of LiClO₄ was added, it may be possible that the spiro ring-opening mechanism did not occur upon the addition of Cu^{2+} . Since the compound itself did not exhibit any fluorescence, the small CHEF effects with Cu^{2+} and Li^+ can be also explained by the blocking of the PET (photoinduced electron transfer) process.

Conclusions

In conclusion, a new fluorescent sensor based on rhodamine B for Pb^{2+} was synthesized. The new fluorescent sensor displayed an extreme selectivity for Pb^{2+} over the other metal ions examined in acetonitrile. An overall emission change of 100-fold was observed upon the addition of Pb^{2+} , and the selectivity for Pb^{2+} was calculated to be 200 times that of Zn^{2+} . Variations in the ligands on the rhodamine B may provide various fluorescent chemosensors, which are selective for different metal ions.

Acknowledgment. This work was supported by the Korean Science and Engineering Foundation (R14-2003-014-01001-0 to J. Y. and Creative Research Initiative Program to W. N.).

Supporting Information Available: NMR spectra, ESI mass spectra and X-ray crystallographic data (CIF) of **1** and its complexes. This material is available free of charge via the Internet at http://pubs.acs.org.

JA051075B